

1 **Archaeophenomics of ancient domestic plants and animals using geometric**
2 **morphometrics : a review**

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21 **Abstract**

22 Geometric morphometrics revolutionized domestication studies through the precise
23 quantification of the phenotype of ancient plant and animal remains. Geometric morphometrics
24 allow for an increasingly detailed understanding of the past agrobiodiversity and our ability to
25 characterize large scale ancient phenotypes has led to what can be named archaeophenomics :
26 the large scale phenotyping of ancient remains. This review describes advances in the
27 bioarchaeological study of domesticated species and their wild relatives where their phenomes
28 are quantified through geometric morphometrics. The two main questions addressed by
29 archaeophenomics are i) taxonomic identification, including domestication signature, and ii)
30 the inference of the spatio-temporal agrobiodiversity dynamics. Archaeophenomics is a
31 growing field in bioarchaeology of domestic species that will benefit in the near future from
32 advances in artificial intelligence and from an increasing interest in multiproxy approaches
33 combining morphometric data with *e.g.* isotopes or archaeogenomics.

34

35 **1. Introduction**

36 Domesticated species have played a major role in the development of Charles Darwin's work,
37 being the subject of the first chapter of the 'Origin of Species' (Darwin, 1859) and later of a
38 dedicated book, 'The variation of animals and plants under domestication'(Darwin, 1868)
39 where he described the mechanisms of variation in domestic species. With his work, Darwin
40 contributed to the understanding of the morphological changes that occurred during the long
41 process of domestication. For most domesticated species, modern breeds and varieties today
42 present a huge morphological diversity reflecting millennia of human selection for many
43 purposes (e.g. food production, work, aestheticism) in various environmental conditions. While
44 the study of the current domestic diversity is mainly carried out in agronomic research, with
45 breed and varietal improvement using molecular breeding programs, a large amount of research
46 has been done to explore the past diversity of domestic species whose remains are found in
47 increasing numbers in archaeological deposits. The methodological development in
48 morphometrics have revolutionized, qualitatively and quantitatively, the study of the phenotype
49 of those remains. Today we have reached a state where the use of several tools, including
50 morphometrics, have allowed phenome (i.e. the full set of observable traits) quantification of a
51 large number of archaeological specimens leading to a renew in the study of archaeological
52 remains of domestic species. We here coin the word *archaeophenomics* for such large-scale
53 quantification of phenotypic data from archaeological specimens. Archaeophenomics, i.e.
54 phenomics of the past, is an emerging field that will likely become a standard for future
55 bioarchaeological studies. This neologism fulfils the needs to express the new realities of
56 bioarchaeological domestication studies.

57 *From phenomics to archaeophenomics*

58 Phenomics, the analysis of high-dimensional phenotypic data, is part of the '-omics' revolution
59 as genomics or proteomics. Phenomics is the new generation of acquisition and analysis of
60 phenotypic data based on techniques which allow a very large amount of quantitative characters
61 to be acquired and processed with minimal handling time. Assessing the full phenome of an
62 organism is illusional (Houle, Govindaraju, & Omholt, 2010) and this is even more true in
63 bioarchaeology. Archaeological remains of plants and animals are often altered by
64 taphonomical processes (e.g. preservation, fragmentation). Despite these inherent constraints
65 associated with studying archaeological material, the large quantity of remains allows for large-
66 scale morphometric analyses.

67 Morphometrics is one of the many tools of *phenomics*. The ‘morphometric revolution’
68 corresponding to the development of *geometric morphometrics* (GMM), *i.e.* the study of forms
69 in multi-dimensional spaces, allow more in-depth investigation of morphological changes
70 (Adams, Rohlf, & Slice, 2004; Rohlf & Marcus, 1993). The main improvement of geometric
71 morphometrics compared to the so-called ‘traditional morphometrics’ is that biological forms
72 are no longer captured by sets of independent measurements of lengths or angles, but by sets of
73 point coordinates, improving dramatically the capture of the geometric complexity of these
74 objects (Bookstein, 1991; Kuhl & Giardina, 1982; Rohlf & Marcus, 1993). Morphometric
75 analyses are often the only available approach for studying the morphology of ancient remains
76 with a fine-scale resolution, while suffering less from preservation limitations than *e.g.* ancient
77 DNA and offering much better possibilities for being carried out on a large scale at a limited
78 cost, in both time and money. This is especially true for plants whose remains are often found
79 charred, a condition strongly detrimental to DNA preservation and which generally prevents
80 the analysis of these remains, at least with current aDNA techniques (Nistelberger, Smith,
81 Wales, Star, & Boessenkool, 2016). Most of the time, **once a specimen is recovered a**
82 **morphometric analysis can be performed**, as long as the structures grabbing the geometrical
83 features analysed are present. While it is always better to analyse complete specimens,
84 fragmented remains can even be studied using a restricted version of the initial protocol since
85 fragmentation does not necessary prevent taxonomical identification (*e.g.* Cornette et al., 2015;
86 Owen et al., 2014, Durocher in press). Geometric morphometric techniques are therefore
87 particularly well adapted to bioarchaeological studies and are a growing field in the discipline.
88

89 Archaeophenomics **appears to be** as a major breakthrough in bioarchaeology, with a drastic
90 quantitative change in the scale of the number of specimens and populations that can be
91 analysed, and a qualitative improvement provided by an increased resolution of the analyses
92 paired with a better description of the morphometric variation with improved detection and
93 visualisation of the shape variation.

94 *Aim and scope of the review*

95 Here we provide an exhaustive review of bioarchaeological studies using archaeophenomics,
96 through geometric morphometrics, to study archaeological remains of domestic species. We
97 restricted our review to studies published in international journals (*i.e.* excluding grey
98 literature), only those focusing on domestic species and that include archaeological specimens.

99 This therefore explicitly excludes: studies of commensal species (*e.g.* rodents (Cucchi, 2008;
100 Cucchi et al., 2014; Valenzuela-Lamas, Baylac, Cucchi, & Vigne, 2011)); studies focusing only
101 on the ancestors and modern relatives of the domestic populations (*e.g.* Late Glacial horse
102 (Bignon, Baylac, Vigne, & Eisenmann, 2005), rabbit (Pelletier, 2019)); and finally the
103 numerous studies focusing only on modern domestic specimens (*e.g.* Battesti et al., 2018;
104 Bonhomme et al., 2017; Evin et al., 2022; Evin, Dobney, et al., 2015; Gros-Balthazard et al.,
105 2016; Hanot et al., 2021; Harbers et al., 2020; Neaux et al., 2020; Pelletier, Kotiaho, Niinimäki,
106 & Salmi, 2020, 2021). Yet these studies are of prime interest for the understanding of ancient
107 populations, they fall out of the scope of this review.

108

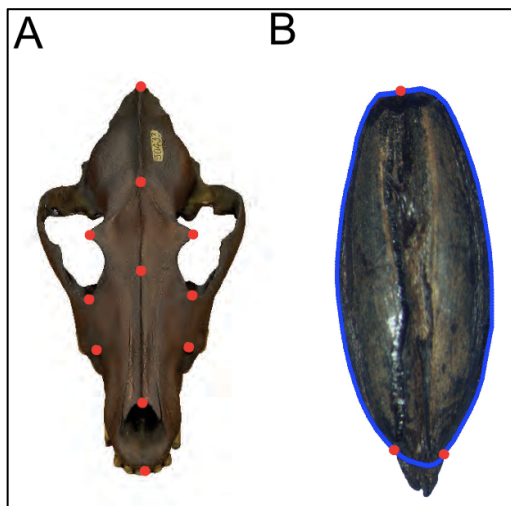
109 2. Geometric morphometrics in archaeophenomics

110 The development of geometric modern morphometrics (GMM) (Bookstein, 1991; Rohlf &
111 Marcus, 1993) came as a response to the conceptual and methodological limits of traditional
112 morphometric methods, such as a better ability to efficiently partition the size and the shape
113 components of the form variation and the possibility of visualizing shape variation. Shape
114 analysis, through geometric morphometrics, allows analysing microscale variation that could
115 not otherwise be identified using traditional techniques. Two main geometric morphometric
116 approaches are currently used in bioarchaeology (fig. 1): **Procrustes approaches** through the
117 acquisition of landmarks and sliding semi-landmark coordinates **and outline analyses** using
118 various methodologies (see below). In bioarchaeology, as in biology, objects are studied in two-
119 or three-dimensions depending on the geometry and size of the remains. Until recently, all
120 archaeobotanical remains appear to have been studied in 2D from digital images, though one
121 recent publication used 3D X-ray-computed tomography to quantify watermelon seeds
122 (Wolcott et al., 2021) (table 1). In addition, archaeobotanical remains are nearly exclusively
123 studied through their outlines geometries using mainly either Elliptic Fourier transforms (EFT)
124 (Giardina & Kuhl, 1977; Kuhl & Giardina, 1982) or natural/orthogonal polynomial equations
125 (Rohlf, 1990) (table 1) and only few studies use landmarks and sliding semi-landmarks
126 coordinates (Ros et al. 2014; García-Granero et al. 2016; Wolcott et al. 2021) (table 1).
127 For animals, teeth are studied in 2D and the same applies to some postcranial bones (*e.g.*
128 phalanges, and talus) although it is quite possible to study them in 3D (Hanot, Guintard, Lepetz,
129 & Cornette, 2017; Haruda, 2017) (table 1). The same applies to mandible that are studied either
130 in 2D (cat: (Vigne et al., 2016), dog: (Ameen et al., 2019)) or 3D (dog: (Drake et al., 2017)).

131 Skulls, which are geometrically more complex objects, are studied in 3D, either directly on the
132 specimens using *e.g.* a Microscribe digitizer (Drake & Klingenberg, 2008; Geiger et al., 2017;
133 Hanot et al., 2017) or through 3D model reconstruction obtained by CT-scanning
134 (Schoenebeck, Hamilton-Dyer, Baxter, Schwarz, & Nussbaumer, 2021) or photogrammetry
135 (dog: (Ameen et al., 2019)). So far, a single study really takes advantage of a CT-scanning
136 technology to analyze the internal structure of the skull that is the inner ear morphometry
137 (Clavel et al., 2021).

138

139 <Figure 1> Example of geometric morphometric protocols applied to bioarchaeological
140 remains. A Example of Procrustes approaches used to quantify the morphometric variation of
141 canid skulls with 3D landmarks. B Example of protocols for 2D outline analyses used to
142 quantify the morphometric variation of barley grains.



143

144

145 3. Domestic species studied with geometric morphometrics

146 We identified a total of 71 studies among which 38 focus on animals and 33 on plants (fig. 2.A,
147 table 1, SI table 1).

148 For animals, only mammals have been studied (though a PhD thesis should be mentioned on
149 chicken (Foster, 2018)), which also represent the large majority of domesticated animals. Ten
150 species are listed and the most represented taxa are pig (N=16), followed by dog (N=9), equids
151 (N=5, horse and donkey), caprines (sheep and goat) (N=5), camelids (N=3, guanaco and llama),
152 and finally cat, and cattle with a single mention (table 1, fig. 2). Animal studies focus primarily
153 on teeth (N=21), skull (N=10, cranium and mandible), in a much larger majority than
154 postcranial bones (*e.g.* talus, phalanges or calcaneus) (SI-table 1).

155 For plants, only angiosperms are concerned and 14 species (or group of closely related species)
156 are listed with grapevine that largely outnumber other species in terms of publications number
157 (N=15), followed by olive (N=6), date palm (N=3) while all other species are only represented
158 by a single mention (table 1). A higher number of studies is dedicated to dicotyledons (N=27)
159 than to monocotyledons (N=6) and focus exclusively on fruits and seeds (SI table 1).

160

161

162 <Table 1> List of the reviewed publications. An extended version of the table, including
 163 research themes, employed methodology, and combination with biomolecular markers can be
 164 found in SI-table 1.

Group	Taxa	References
Mammals	Pig	(Balasse et al., 2016; Bartosiewicz et al., 2013; Bopp-Ito, Cucchi, Evin, Stopp, & Schibler, 2018; Cucchi et al., 2016, 2021; Cucchi, Fujita, & Dobney, 2008; Cucchi, Hulme-Beaman, Yuan, & Dobney, 2011; Dobney, Cucchi, & Larson, 2008; Duval, Cucchi, Horard-Herbin, & Lepetz, 2018; Duval, Lepetz, Horard-Herbin, & Cucchi, 2015; Evin, Flink, et al., 2015; Frémondeau, De Cupere, Evin, & Van Neer, 2017; Krause-Kyora et al., 2013; Marom et al., 2019; Ottoni et al., 2013; Price & Evin, 2017)
	Dog	(Ameen et al., 2019; Daza Perea, 2017; Drake, Coquerelle, & Colombeau, 2015; Drake et al., 2017; Drake & Klingenberg, 2008; Fisher, 2019; Geiger et al., 2017; Manin & Evin, 2020; Schoenebeck et al., 2021)
	Caprines (sheep and goat)	(Colominas et al., 2019; Haruda, 2017; Haruda, Varfolomeev, Goriachev, Yermolayeva, & Outram, 2019; Pöllath, Alibert, Schafberg, & Peters, 2018; Pöllath, Schafberg, & Peters, 2019)
	Equids (horse and donkey)	(Chuang & Bonhomme, 2019; Clavel et al., 2021; Cucchi et al., 2017; Hanot et al., 2017; Seetah, Cardini, & Barker, 2016)
	Camelids (guanaco and llama)	(Hernández, L'Heureux, & Leoni, 2021)
	Cat	(Vigne et al., 2016)
	Cattle	(Cucchi et al., 2019)
Monocotyledons	Date palm	(Gros-Balthazard et al., 2017; Sallon et al., 2020; Terral et al., 2012)
	Barley	(Jérôme Ros, Evin, Bouby, & Ruas, 2014)
	Millet	(García-Granero et al., 2016)
	Wheat	(Bonhomme et al., 2016)
Dicotyledons	Lemon	(Grasso, Mavelli, & Fiorentino, 2018)
	Melon	(Sabato et al., 2019)
	Grapevine	(Bacilieri et al., 2017; Bonhomme et al., 2020; Bonhomme, Terral, et al., 2021; Bouby et al., 2018, 2021; Figueiral et al., 2015; Mariotti Lippi et al., 2020; Margaritis et al., 2021; Moricca et al., 2021; Orrù, Grillo, Lovicu, Venora, & Bacchetta, 2013; Pagnoux et al., 2015, 2021; Terral et al., 2010; Uccesu et al., 2015, 2016; Valamoti et al., 2020)
	Olive	(Bourgeon et al., 2018; Margaritis et al., 2021; Newton, Lorre, Sauvage, Ivorra, & Terral, 2014; Newton, Terral, & Ivorra, 2006; Terral et al., 2004, 2021)
	Opium poppy	(Jesus et al., 2021)
	Cherry	(Burger, Terral, Ruas, Ivorra, & Picq, 2011)
	Pulses (grass pea, lentil, broad bean)	(Tarongi et al., 2020)
	Watermelon	(Wolcott et al., 2021)

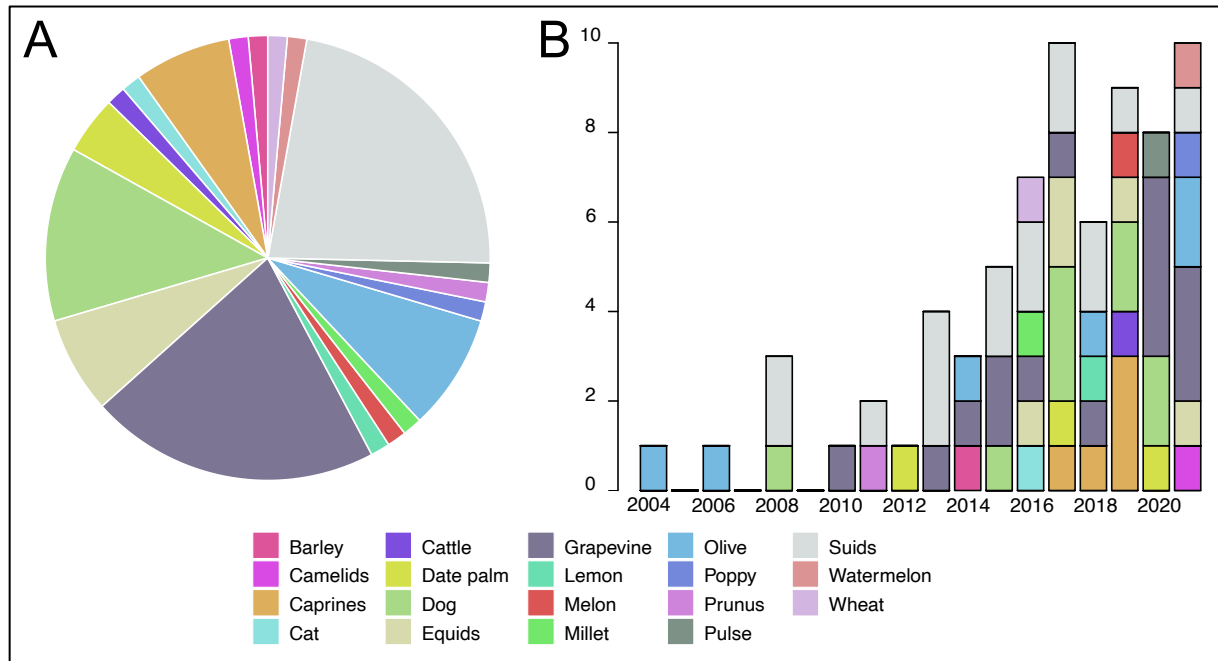
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166 Since 2004, year of the first publication included in this review (Terral et al., 2004), the yearly
 167 number of published bioarchaeological studies on domestic animal and plant species using
 168 geometric morphometrics is steadily increasing (Fig. 1.B).

169

170

171 <Figure 2> Bioarchaeological archaeophenomic studies, concerning domestic species and
 172 using geometric morphometrics, represented by taxa (A) and year (B). A: relative frequencies
 173 of the different groups studied. B: Evolution of the number of studies per year. Details of the
 174 references can be found in table 1 and SI table 1.



175

176 The number of phenomes now available greatly differs between species with grapevine coming
 177 out on top with a maximum of 2223 archaeological seeds having been quantified in one single
 178 study (Pagnoux et al., 2021). The number of papers published per year is still low compared to
 179 *e.g.* palaeogenomic studies (~1480 references obtained for a quick online search of the term
 180 “palaeogenomics” in google scholar the 11/02/2022). Numbers of studied individuals are
 181 usually lower for animals which are mostly represented by fewer individuals per archaeological
 182 assemblage. It should be noted, however, that the concept of ‘individual’ differs here between
 183 archaeozoology and archaeobotany, since an animal will be represented by single elements such
 184 as a cranium or a lower right third molar, while numerous studied individuals (*e.g.* seeds or
 185 fruit-stones) may come from a single plant individual.

186 4. Main bioarchaeological research themes based on GMM data

187 Archaeophenomics through geometric morphometrics is increasingly used for bio-
 188 archaeological studies for two main purposes that are the **taxonomic identification** of the
 189 archaeological remains including the domestication signature, and to assess the
 190 **agrobiodiversity variation in both time and space** primarily related to processes of

191 colonisation-dispersal, adaptation, diversification and changes in husbandry or cultivation
192 practices or cultural choices.

193

194 ***Taxonomic identification and domestication signature***

195 A prerequisite of many bioarchaeological studies is to perform a taxonomical identification of
196 the remains, either by identifying the taxa to which the specimen belongs and/or to identify its
197 wild or domestic status. Archaeological remains are often fragmented or altered due to either
198 taphonomic due to various anthropic and taphonomic processes (*e.g.* charring, butchery or
199 culinary preparation) processes rendering their identification potentially delicate. Geometric
200 morphometric protocols are now available to distinguish (more or less effectively depending
201 on the study and model) morphologically close mammalian species of equids, bovids, sheep-
202 goats, camelids, canids, cats and pigs (table 1). Taxonomic identification of plant remains is
203 even more challenging due to the larger number of species (or sub-species) potentially
204 occurring at an archaeological site. Botanical studies dealing with taxonomic identification
205 include cereals, such as barley, millet, einkorn and emmer, opium poppy, pulses, citrus, melon,
206 watermelon, date palm and prunus species (SI table 1). For other remains whose species
207 identification is unambiguous, the question of the wild and domestic status distinction and
208 identification may arise. This is especially true for species that have a wild ancestor with a wide
209 geographical range and for which the wild and domestic populations have coexisted for
210 millennia. This is the case for nearly all species with the exception of those having an ancestor
211 leaving in a restricted geographic area (*e.g.* sheep, goat and most cereals). This geographic
212 proximity can also be source of hybridization between wild and domestic individuals as already
213 documented from genomic and palaeogenomic data. for *e.g.* pig (Frantz et al., 2019), dog (Pilot
214 et al., 2018), grapevine (Myles et al., 2011; Riaz et al., 2018) or date palm (Gros-Balthazard et
215 al., 2017) or among individuals of distinct species as evidenced for north African date palm
216 (Flowers et al., 2019) which can render their morphometric identification even more
217 challenging. In addition, studies looking at bridging archaeological samples to modern breeds
218 or varieties, or groups of them, are more often found for perennial clonal plants, that show an
219 extended varietal diversity and for which varieties can theoretically persist substantially
220 unchanged for centuries or even millennia through vegetative multiplication (*e.g.* grapevine,
221 olive, *Prunus* species and date palm (table 1)). For non-perennial clonal organisms, such as
222 animals, it seems that direct comparison to specific modern breeds has been done, so far, only
223 for dogs (Geiger et al., 2017; Schoenebeck et al., 2021), and that such comparison can be

224 questioned as the intensification of selective pressures during the last centuries likely
225 dramatically altered ancient morphologies.

226 Another specificity of archaeobotanical remains compared to zooarchaeological ones is that
227 remains are often found charred and that charring can affect their size and shape and therefore
228 their taxonomic identification. An important effort has been made for multiple taxa to
229 understand the effect of charring on the morphometric results and their interpretations (*e.g.*
230 cereals (Bonhomme et al., 2017; Ros et al., 2014), grapevine (Bouby et al., 2018; Uccesu et
231 al., 2016), and olive (Terral et al., 2004)).

232 A significant proportion of the domestic species whose remains are found during archaeological
233 excavations have been the subject of geometric morphometrics and such approaches have been
234 found effective for taxonomic identification of the remains. The many available protocols can
235 now be adapted to nearby species not yet subjected to such studies.

236

237 ***Documenting spatio-temporal variation of past agrobiodiversity***

238 A large number of archaeophenomic studies explore the morphometric spatio-temporal
239 variation of domestic populations. Such studies span either long periods of time of several
240 millennia (*e.g.* Pagnoux et al., 2021; Price & Evin, 2017; Terral et al., 2004), or a much shorter
241 period of no more than a century (*e.g.* Drake & Klingenberg, 2008).

242 Time and space are intertwined components of bioarchaeological studies. It is however possible
243 to study them separately by comparing either diachronic populations of a single locality or
244 synchronous populations of various geographic origins.

245 The studies that explore the geographic variation between populations from a single chrono-
246 cultural period (SI-table 1) evidenced that both geographically near and far populations can
247 show morphometric differences. In term of interpretations, a geographic structuration of
248 synchronous populations may correspond not exclusively to local environmental adaptation,
249 different husbandry/cultivation practices, cultural choices or distinct genetic lineages.

250 Similarly, diachronic differences between populations originating from the same geographic
251 area can reveal either changes in human practices, spread of new genetic stock, environmental
252 variation, or drift. Such comparison between diachronic populations (SI-table 1) can reveal both
253 long term variation, but also more abrupt morphological shift between periods.

254 In comparison, fewer studies focus both on time and space (SI-Table 1). Generally, while
255 absence of differences between assemblages cannot lead to the conclusion that they belong to
256 similar populations, on the other hand, the existence of differences allows to hypothesize the
257 existence of distinct populations with at least limited cultural or genetic exchanges. As a

258 consequence, morphometric analyses can make only limited contribution to mobility studies,
259 that can be better explored using *e.g.* biomolecular markers such as ancient DNA or isotopes.
260 In the same way, phenotypic proximity does not necessarily reflect genetic proximity (*i.e.*
261 phylogeny) due to natural or anthropic selection. During domestication, human populations
262 have selected certain traits such as larger quantity of meat or of fruit size. In archaeophenomics
263 studies, the target of human selection during domestication and subsequent diversification is
264 not necessarily the target of the morphometric analysis. For cereals, grain size was likely
265 intentionally selected, but not their shape, while for fruit stones neither size or shape were likely
266 directly targeted, even if in some cases (*e.g.* at least for grapevine) seed and fruit measurements
267 covariate (Bonhomme et al., 2020). Similarly, in mammals, it is unlikely that teeth, that are
268 commonly studied and considered as a phenotypic marker of adaptation to natural or anthropic
269 environment, were not likely the aim of human selection that primarily targeted primary (*e.g.*
270 meat) or secondary products (*e.g.* milk, wool). In all these cases, where domestic and wild
271 populations differ in size and shape of anatomical elements non targeted by selection, other
272 evolutionary pressures and mechanisms such as drift, genetic hitchhiking or indirect selection
273 (*e.g.* morpho-functional constraint) can be invoked. In addition, many anatomical structures are
274 polygenic (*e.g.* Harjunmaa et al., 2012), or the genes involved are not known. Moreover, not all
275 anatomical elements necessarily evolve in parallel, at the same rate or following the same
276 direction (*e.g.* Geiger & Sánchez-Villagra, 2018).

277

278 **5. Multi-proxy approaches and future methodological developments**

279 Archaeobotanical remains are often conserved through charring which is detrimental to DNA
280 preservation (Nistelberger et al., 2016) rendering the combination of such approaches with
281 morphometric data impossible. This is however possible to the less frequently found
282 waterlogged remains where DNA can be preserved and the results compared to morphometric
283 data (Bacilieri et al., 2017; Bouby et al., 2021).

284 For animals, several studies combined geometric morphometrics with ancient DNA (*e.g.* pig:
285 (Evin, Flink, et al. 2015), dog (Ameen et al., 2019)), geometric morphometrics and isotopes
286 (pig: (Cucchi et al., 2016; Frémondeau et al., 2017)), or the combination of the three approaches
287 geometric morphometrics, ancient DNA and isotopes (pig: (Balasse et al., 2016)). Isotopic
288 analyses are increasingly used in archaeobotany (*e.g.* Fiorentino, Ferrio, Bogaard, Araus, &
289 Riehl, 2015), but not yet in combination with other approaches.

290 Artificial intelligence is increasingly used in biology (*e.g.* Ching et al., 2018; Hassoun et al.,
291 2021) and archaeology (*e.g.* Bickler, 2021; Horn et al., 2021), but only few bioarchaeological

292 studies use yet such approaches (*e.g.* Miele, Dussert, Cucchi, & Renaud, 2020). Machine
293 learning in general, and deep learning using convolutional neural networks in particular, will
294 certainly help in the future for automatic data acquisition such as landmark coordinates (*e.g.*
295 Devine et al., 2020), image post-treatment prior to outline analyses, and/or directly for binary
296 (status) or multiple (species identification) classification tasks. In addition, the ever-growing
297 motivation to share data and knowledge should drastically extent the chrono-cultural and
298 geographic scopes of the studies allowing comparisons not possible before. This would be
299 possible only after careful inter-operator and methodological comparisons (Evin, Bonhomme,
300 and Claude 2021). Other future lines of research will also certainly focus on evo-devo
301 perspectives (Bonhomme et al., 2020), form-function interactions (Harbers et al., 2020; Neaux
302 et al., 2020) as well as further exploration of the genotype-phenotype relationships
303 (Schoenebeck & Ostrander, 2013).

304

305 **6. Conclusion/perspectives**

306 Archaeophenomics through geometric morphometrics allows addressing questions regarding
307 the micro-evolutionary processes that accompanied the long history of domestic species in an
308 unprecedented way. Such approaches are increasingly used in bioarchaeology and are
309 becoming one of the many approaches now available to us to study past populations. The future
310 of phenotypic studies will require carefully thought, managed and open large-scale databases,
311 precisely contextualised archaeologically, and combining the whole set of available
312 approaches, if possible carried out on the exact same specimens. As for the relatively recent
313 research fields of palaeoproteomics or palaeogenomics, this review shows that
314 archaeophenomics definitely corresponds to a renew of domestication studies deserving a new
315 terminology. This review **attempts** to list all the studies in the scope of archaeophenomics where
316 the phenomes of domestic species are quantified through geometric morphometrics. **The many**
317 **approaches now available pave the way to future research expanding the diversity of studied**
318 **species and the archaeological questions that can be addressed through archaeophenomics.**

319

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330

331 **Conflict of interest disclosure**

332 The authors declare they have no conflict of interest relating to the content of this article.

333

334 **Supplementary information**

335 SI-Table 1 is available in the open repository OSF with the DOI : 10.17605/OSF.IO/T3Q96.

336

337

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